

# Efficacy of a plant-produced infectious bronchitis virus-like particle vaccine in specific pathogen-free chickens

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**ABSTRACT** Infectious bronchitis (IB) Gammacoronavirus causes a highly contagious respiratory disease in chickens that is listed by the World Organisation for Animal Health (WOAH). Its high mutation ability has resulted in numerous variants against which the commercially available live or recombinant vaccines singly offer limited protection. *Agrobacterium*-mediated transient expression in *Nicotiana benthamiana* (tobacco) plants was used here to produce a virus-like particle (VLP) vaccine expressing a modified full-length IBV spike (S) protein of a QX-like IB variant. In a challenge study with the homologous live IB QX-like virus, VLP-

vaccinated birds produced S protein-specific antibodies comparable to those produced by live-vaccinated birds seroconverting with mean geometric titers of 6.8 and 7.2 log<sub>2</sub>, respectively. The VLP-vaccinated birds had reduced oropharyngeal and cloacal viral shedding compared to an unvaccinated challenged control and were more protected against tracheal ciliostasis than the live-vaccinated birds. While the results appeared similar, plant-produced IB VLPs are safer, more affordable, easier to produce and update to antigenically match any emerging IB variant, making them a more suitable alternative to IBV control than live-attenuated vaccines.

**Key words:** *N. benthamiana*, infectious bronchitis, vaccine, virus-like particle

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## INTRODUCTION

Infectious bronchitis virus (IBV), listed by the World Organisation for Animal Health (WOAH) as a notifiable disease, causes extremely transmissible respiratory disease in chickens (*Gallus gallus*). IBV has an extensive economic impact on the global poultry industry, second only to that of avian influenza (World Bank & TAFS-Forum, 2011).

IBV is classified as a *Gammacoronavirus* of the *Coronaviridae* family, a single-stranded, enveloped RNA virus with a pleomorphic or sphere-shaped form, surrounded by club-shaped spikes (Jackwood and de Wit, 2013). The IBV spike (S) protein induces neutralizing antibodies (Kant et al., 1992; Jackwood and de Wit, 2013), but mutations occurring in the S protein S1 subunit result in the emergence of novel IBV serotypes that can evade protection offered by conventional serotype-specific vaccine types, such as live-attenuated or inactivated vaccines (Shirvani et al., 2018; Gallardo, 2021). At least 7 IBV

genotypes exist, with over 35 different serotypes offering little or no cross-protection against antigenically distinct variants (Valastro et al., 2016), making IBV control exceedingly challenging with commercial serotype-specific vaccines (de Wit et al., 2011). However, in some cases the so-called “protectotype” phenomenon is useful, whereby combining 2 antigenically distinct IB vaccine serotypes confers broad serologic cross-protection against a third, unrelated serotype (de Wit et al., 2011; Butcher et al., 2022). For example, a combination of 4-91 and Mass-type variant vaccine strains is commonly used which has been shown to increase the level of protection against IBV in young chickens following challenge with different IBV variants (Terregino et al., 2008; Valastro et al., 2016). Elevated levels of protection can be achieved by priming with a live-attenuated vaccine prior to boosting with an inactivated vaccine or better yet, an inactivated autogenous variant vaccine (de Wit et al., 2011; Erfanmanesh et al., 2020). However, the production of autogenous variant vaccines normally entails the isolation and propagation of field virus in embryonated chicken eggs (Vaughn and Whitehead, 2009).

Virus-like particles (VLP) resemble native viruses, but lack the genetic material required to facilitate replication and cause infection (Ukarami et al., 2017). Several adaptations of VLPs against IBV have been

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developed in baculovirus expression vector systems, that could induce strong humoral and cellular immune responses in chickens, comparable to those elicited by inactivated IBV vaccines (Liu et al., 2013; Lv et al., 2014; Xu et al., 2016; Wu et al., 2019). Previously, a recombinant plant-produced IB VLP vaccine was successfully developed, with a modified full-length IBV S protein containing stabilizing proline mutations (Pallese et al., 2017), and the following modifications: 1) the transmembrane domain (TM) and cytosolic tail (CT) of IBV substituted with that of a Newcastle disease virus (NDV) fusion protein (F), and 2) a murine signal peptide replacing the native signal peptide, coexpressed with the NDV matrix (M) protein (Sepotokele et al., 2023). A single immunization with the IB VLP vaccine was able to induce robust immune responses in specific pathogen-free (SPF) chickens after 2 wk (Sepotokele et al., 2023).

The present study further explored the protective efficacy of this IB VLP vaccine in SPF White Leghorn chickens, in a prime-boost strategy with live commercial vaccines that are known to confer protection against the QX-like variant used for challenge.

## MATERIALS AND METHODS

### Plant-Produced IB VLP Vaccine Production

The AGL-1 *Agrobacterium* stock containing the recombinant pEAQ-HT plasmid for the modified construct designated mIBV-S2P-NDV-F<sup>TM/CT</sup> (Sepotokele et al., 2023) was propagated overnight at 28°C in Luria broth containing the selective antibiotics to an OD<sub>600</sub> of ≤2. The recombinant plasmid for the NDV M protein, which had already been transformed into AGL-1, was propagated in the same way.

The overnight cultures were centrifuged at 10°C for 7 min at 7,000 × *g*, the pellet was resuspended in MES (2-(N-morpholino)ethanesulfonic acid) buffer, and diluted to an OD<sub>600</sub> of between 1 and 1.5. The pEAQ-HT-mIBV-S2P-NDV-F<sup>TM/CT</sup> construct was combined to a 4:1 ratio with pEAQ-HT-NDV-Matrix, and the mixture was left for 1 h at room temperature prior to infiltration. It was previously demonstrated that a ratio of 4:1 elevated the expression and assembly of IB VLPs (Sepotokele et al., 2023).

The leaves of 3- to 4-wk-old *Nicotiana benthamiana* ΔXT/FT plants (Strasser et al., 2008) were infiltrated with the prepared *Agrobacterium* culture combination using a needle-less syringe. Six days after agro-infiltration, the leaves were harvested, weighed and homogenized in a juicer in 2 volumes of either 1× Tris buffer (50 mM Tris base, 150 mM NaCl, pH 8.0), 1× Bicine buffer (20 mM NaCl, and 50 mM Bicine; pH 8.4) or 1× phosphate buffered saline (PBS) (140 mM NaCl, 1.5 mM KH<sub>2</sub>PO<sub>4</sub>, 10 mM Na<sub>2</sub>HPO<sub>4</sub>, 2.7 mM KCl, pH 7.4) containing protease inhibitor cocktail (Sigma-Aldrich, St. Louis, MO), and 0.04% sodium metabisulfite. The homogenized leaf extract was purified using

sucrose density ultracentrifugation as previously described (Sepotokele et al., 2023).

Fractions 3 to 6 of the collected sucrose density gradient fractions were separated on a 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gel and stained with Coomassie stain overnight. Western blots were performed with the same fractions using a 1:1,000 dilution of IBV antisera from commercial birds that had been vaccinated with live-attenuated Mass-type commercial vaccines. The secondary antibody used was a 1:2,000 dilution of Goat-α-Chicken IgY HRP (Abcam, Novex). Clarity Western ECL chemiluminescence substrate (BioRad, Hercules, CA) was used for protein detection on the ChemiDoc MP Imaging System (BioRad).

Carbon-coated holey copper grids of the plant-produced IB VLPs and SPF chicken egg allantoic fluid (AF) containing a live propagated IBV (strain ck/ZA/3665/11) were prepared as described previously (Sepotokele et al., 2023) and were imaged using a flash transmission electron microscope (TEM), JEOL JEM-1400, at the University of Pretoria.

The fractions containing the highest level of S protein expression as confirmed by SDS-PAGE and Western blot were pooled (i.e., those for mIBV-S2P-NDV-F<sup>TM/CT</sup>:Matrix processed in PBS buffer), dialyzed in 1× PBS using 3500 mW CO Slide-A-Lyzer Dialysis Cassettes (ThermoFisher Scientific, Waltham, MA), and stabilized with 15% (w/v) trehalose dehydrate (Sigma-Aldrich). The fractions containing the IB VLPs harvested in Tris and Bicine buffers were also dialyzed for comparison.

The partially purified VLPs were quantified using densitometric analysis. Bovine serum albumin (BSA) protein standards in known concentrations were electrophoretically separated alongside the dialyzed VLPs on a 12% SDS-PAGE gel. The overnight stained gel was analyzed using the ChemiDoc MP Imaging System Quantification Software (BioRad) with molecular weight markers and the protein standards serving as references.

### Vaccines and Challenge Virus Preparation

Live freeze-dried IB 4-91 and IB Ma5 vaccines (both sourced from Nobilis, MSD Animal Health, South Africa) were reconstituted separately in Diluvac (10 mL) (Nobilis, MSD Animal Health, South Africa) to the recommended single doses of ≥10<sup>3.6</sup> and >10<sup>3.5</sup> EID<sub>50</sub>, respectively; thereafter 1 mL of each was combined into 10 mL of Oculo Nasal Diluent (Nobilis, MSD Animal Health, Abnova, Kempton Park, South Africa).

The dialyzed VLPs were diluted to 20 μg (S protein content) per 225 μL with 1× PBS, and 10% (v/v) of EmulsigenP Adjuvant (MVP, Phibro Animal Health, IDEXX Laboratories Inc.: Omaha, NE) was added to a 250 μL volume.

Challenge virus strain ck/ZA/3665/11 (Bwala et al., 2018) was propagated in SPF chicken eggs and diluted

in Diluent Oculo Nasal to an  $EID_{50}$  of  $10^6$  per  $60 \mu\text{L}$ , that is, 1 drop ( $30 \mu\text{L}$ ) per eye of each bird. The vaccines and challenge virus were prepared on the day of use and kept at  $4^\circ\text{C}$  until administration.

## Animals

The vaccine efficacy challenge study was performed in isolators at the University of Pretoria's Biosafety Level 3 (BSL3). Three-week-old White Leghorn SPF chickens ( $n = 40$ ; Avi-Farms (Pty.) Ltd., Pretoria, South Africa) were numbered individually with neck tags, and assigned randomly into 4 treatment groups, with 10 birds per isolator as follows: Group A: Live + VLP vaccinated; Group B: Live + Live vaccinated; Group C: Unvaccinated challenged control; Group D: Unvaccinated unchallenged control. Water and feed (Nova Feeds, Pretoria, South Africa) were provided ad libitum for the duration of the trial.

## Experimental Design

At d 0, 1 mL blood samples were taken from the wing veins of each bird prior to vaccination. Groups A and B were immunized with a single drop of the prepared live vaccine in each eye. Blood samples were collected again on d 21 from the birds in Groups A and B, before immunizing. Group B was again immunized via the intraocular route with another dose of mixed live vaccine, whereas the birds in Group A were immunized via inoculation in the breast with the  $250 \mu\text{L}$  dose of the prepared VLP vaccine. On d 42, blood samples were collected from Groups A and B, prior to challenging. Groups A, B, and C were challenged via the intraocular route with the live QX-like virus. The birds were monitored for adverse vaccine effects daily throughout the trial, and for clinical signs postchallenge. Oropharyngeal and cloacal swabs were collected from all birds (Groups A, B, C, and D) on d 3, 5, and 7 post-challenge (dpc) using sterile rayon-tipped swabs with plastic applicators (Copan Diagnostics Inc., Murrieta, CA). The swabs were individually placed in 2 mL of viral transport media (0.1 mg/mL doxycycline, 1 mg/mL penicillin-streptomycin, brain-heart broth, 10% glycerol, 0.1 mg/mL enrofloxacin) and stored at  $4^\circ\text{C}$  prior to processing. Blood samples were taken again on d 49 (7 dpc) before euthanasia by cervical dislocation. The tracheas of each bird were removed directly after euthanasia, and placed into PBS for immediate ciliary motility scoring. The challenge model for strain ck/ZA/3665/11 whereby tracheas must be collected at d 7 pc for optimal vaccine efficacy evaluation was previously determined (Bwala et al., 2018).

## Serological Tests

Sera were harvested from clotted blood by low-speed centrifugation ( $3,000 \times g$  at  $22^\circ\text{C}$  for 10 min)

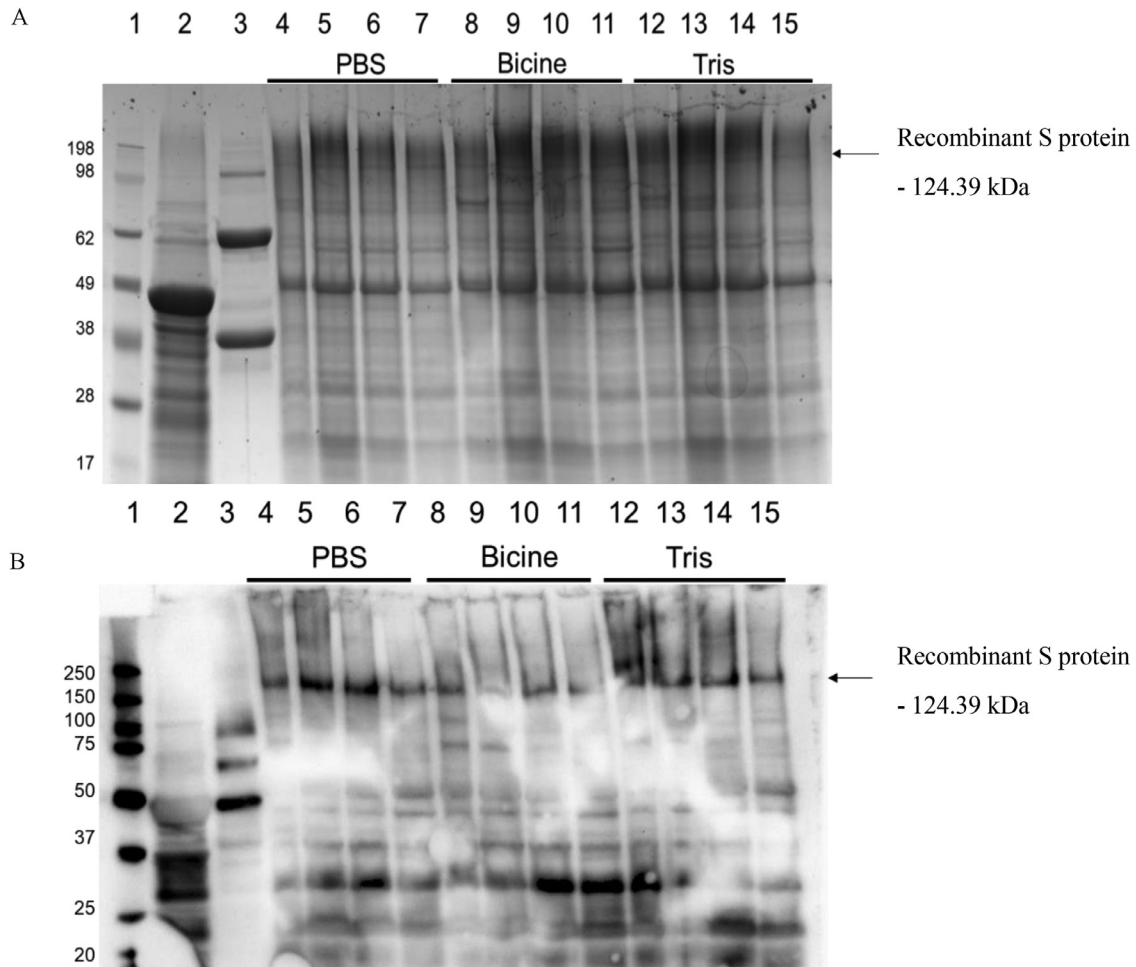
and tested using the commercial IDEXX IBV Antibody Kit (IDEXX Laboratories Inc.) following manufacturer's instructions. For the hemagglutination inhibition tests (HI), antigen was prepared by centrifuging AF from SPF eggs in which strain ck/ZA/3665/11 had been propagated to a titer of  $10^{6.5}$   $EID_{50}/0.1 \text{ mL}$ , for 15 min at  $4^\circ\text{C}$ . Five milliliters of the clarified AF was treated with 1 Unit of neuraminidase from *Clostridium perfringens* (P5290, Abnova, Taiwan) and incubated at  $4^\circ\text{C}$  overnight (Ruano et al., 2000). The HI tests were performed with 1% (v/v) chicken red blood cells (CRBC) as per the recommended method of the WOAAH (WOAH, 2019), but at room temperature. The last well in which CRBC streaming was observed in a 2-fold titration of the test serum relative to the controls was recorded as the  $\log_2$  HI titer.

## Quantification of QX-Like IBV

Total nucleic acid was extracted from 0.2 mL of the swab fluids using the IndiMag Pathogen Kits as recommended in the IndiMag 48 instrument (Indical BioSciences, Leipzig, Sachsen, Germany). The presence of QX-like IBV-specific RNA was detected using the Vetmax-Plus One Step RT-PCR kit (Applied Biosystems, Waltham, MA) in a StepOnePlus instrument (Applied Biosystems) as described in Bwala et al. (2018). This assay was designed and shown to specifically detect only the S-protein gene sequence of the QX-like variant, but not any other serotypes, including the 4-91 and Ma5 vaccine strains. The qRT-PCR reactions included  $0.15 \mu\text{L}$  of the QX-like-specific IBV MGB-FAM-labeled probe ( $5 \text{ pmol}/\mu\text{L}$ ),  $4 \mu\text{L}$  of extracted RNA,  $6 \mu\text{L}$  of  $2\times$  RT-PCR buffer,  $0.5 \mu\text{L}$  each of IBV reverse and forward primer ( $12.5 \text{ pmol}/\mu\text{L}$ ),  $0.5 \mu\text{L}$  of  $25\times$  RT-PCR enzyme mix, and nuclease-free water to make the volume up to  $13 \mu\text{L}$ . The RT-PCR cycling profile was as follows: 10 min at  $48^\circ\text{C}$ , 10 min at  $95^\circ\text{C}$ , and forty cycles of 15 s at  $95^\circ\text{C}$  plus 45 s at  $60^\circ\text{C}$ . Test samples were run alongside a standard curve prepared from the titrated ck/ZA/3665/11 stock and no-template negative controls. The limit of detection of the qRT-PCR assay was  $<15.93 EID_{50}$  equivalents/mL (results of this study).

## Ciliary Motility Scoring

Immediately after the birds were euthanized, their tracheas were removed and submerged in 10 mL of  $1\times$  PBS in 15 mL Universal tubes. The tracheas were cut into 10 ring sections, 3 from the upper, 4 from the mid-section and 3 from the lower section, floated in drops of  $1\times$  PBS in a demarcated Petri dish, and analyzed for ciliary motility across the circumference of the tracheal section under a light microscope at  $10\times$  magnification. Each of the 10 rings for each bird was given a percentage correlating to a score ranging from 0 to 4 as follows: 0:



**Figure 1.** SDS-PAGE (A) and Western blot (B) of plant-produced spike protein (synthetic construct mIBV-S2P-NDV-F<sup>TM/CT</sup>) coinfiltrated at a 4:1 ratio with the NDV matrix protein. Lane 1: molecular weight marker; Lane 2: plant-expressed empty pEAQ-HT vector; Lane 3: purified live QX-like IBV strain ck/ZA/3665/11; Lanes 4 to 7: Fractions 3 to 6 extracted in PBS buffer; Lanes 8 to 11: Fractions 3 to 6 extracted in Bicine buffer; Lanes 12 to 15: Fractions 3 to 6 extracted in Tris buffer.

75 to 100% motility (normal); 1: 51 to 75% motility; 2: 30 to 50% motility; 3: 3 to 30% motility; 4: 0 to 2% motility (little to no motility). The average protection score for each group was calculated using the following equation (Andrade et al., 1982):

Average Protection Score

$$= 100 - \frac{\text{sum total of individual scores in group}}{\text{total number of individuals within group} \times 20} \times 100$$

### Statistical Analysis

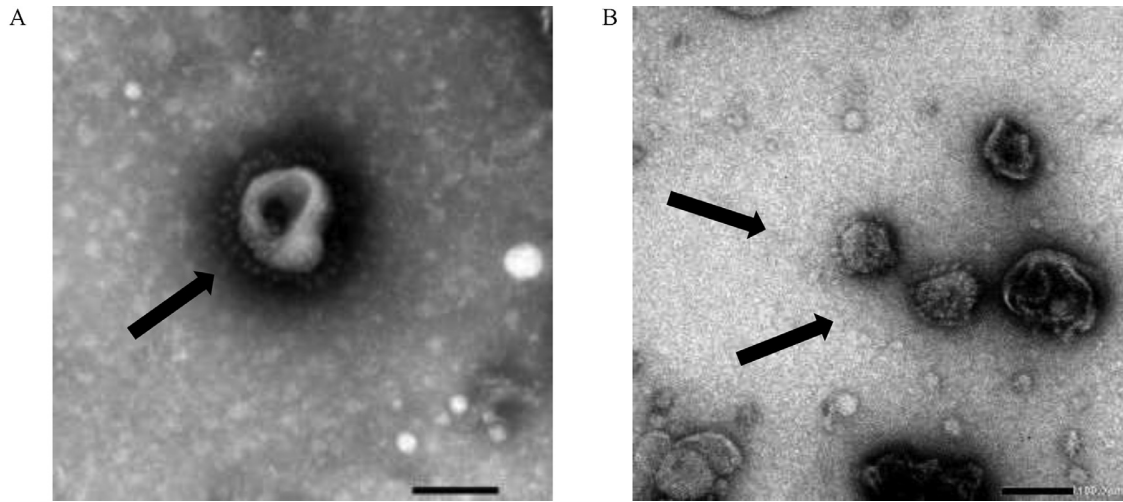
GraphPad Prism v 9.4.1 software for Windows (La Jolla, CA) was used for statistical analysis. The results for the HI tests were compared statistically by means of the unpaired *T* test and a *P* value less than 0.05 was considered significant. The results of the qRT-PCR reactions were analyzed by means of 1-way analysis of variance (ANOVA) followed by the Tukey's multiple comparisons test. The tests were performed on the untransformed data, with the results presented as log<sub>10</sub> values for ease of comparison.

## RESULTS

### Production of the VLP Vaccine in Plants

The leaves of agroinfiltrated tobacco plants were harvested after 6 d, and the IB VLP yields obtained through purification in PBS, Tris, or Bicine buffer were compared. Analysis by SDS-PAGE and immunoblot with IBV antisera confirmed S protein expression with a 124 kDa band correlating to the expected size (Figure 1). The immunoblot showed stronger S protein-specific detection with the antisera in the samples where PBS was used for extraction than in the samples where either Bicine or Tris was used (Figure 1B). The QX-like antisera did not detect a band correlating to the size of the S protein in the positive control (purified live virus), although the flock from which the antisera was collected had been immunized with IBV vaccines of the Mass serotype only, and the level of cross-protection seen between single serotypes is typically low (Cavanagh, 2003).

IB VLPs extracted in either Bicine or Tris buffer yielded no VLPs when analyzed by TEM (data not shown), while the sample harvested with PBS contained VLPs that resembled typical IBV particles in



**Figure 2.** Negative-stained transmission electron microscopy images of (A) live QX-like IB virus strain ck/ZA/3665/11 and (B) plant-produced IBV virus-like particles displaying the spike protein expressed with the synthetic gene construct mIBV-S2P-NDV-F<sup>TM/CT</sup> in *N. benthamiana* in different ratios with NDV matrix. Arrows indicate VLPs.

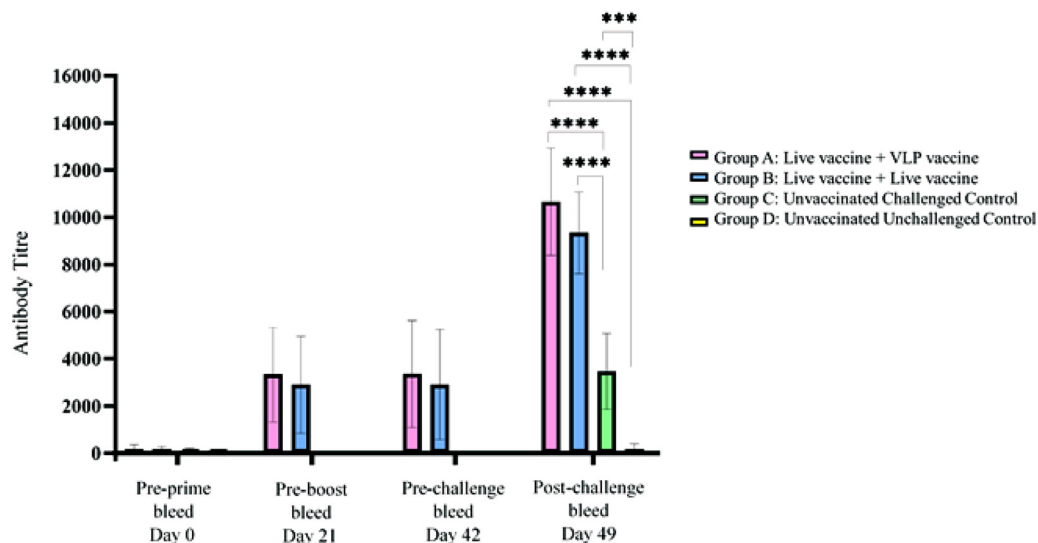
morphology and size, ranging from 67 to 135 nm in diameter (Figure 2). The dialyzed samples from all 3 extraction methods were analyzed using densitometry, but the S protein was undetectable in the samples extracted with either Bicine or Tris, while the sample extracted in PBS had an S protein concentration of approximately 67 ng/ $\mu$ L for the S protein band. The latter sample was therefore used to prepare the vaccine in the efficacy study. Unlike plant-produced avian influenza VLPs (Smith et al., 2020), hemagglutination (HA) could not be used to quantify the IB VLPs because they do not agglutinate chicken red blood cells (Sepotokele et al., 2023).

## Efficacy Study

**Clinical Signs** There were no observable clinical signs in any of the birds, including the nonvaccinated

challenged controls in group C, but the lack of clinical signs was consistent with previous results in SPF chickens infected with strain ck/ZA/3665/11 (Bwala et al., 2018). No adverse vaccine effects were reported for any of the birds following immunization throughout the trial.

**Immunogenicity in Chickens of IB Vaccines Pre- and Postchallenge** As expected, no IB-specific antibodies were detected prior to vaccination (Table S1), and after 21 d, birds in Groups A and B immunized with a mix of the live 4-91/Mass type vaccines, tested positive for the presence of IBV antibodies with average antibody titers of 3347 and 2903, respectively. Twenty-one days later, after birds were boosted with the VLP vaccine (Group A) or with the live vaccine mix (Group B), only marginal increases in the IBV-specific antibodies were detected, with average antibody titers of 3354.49 and 2911, respectively.

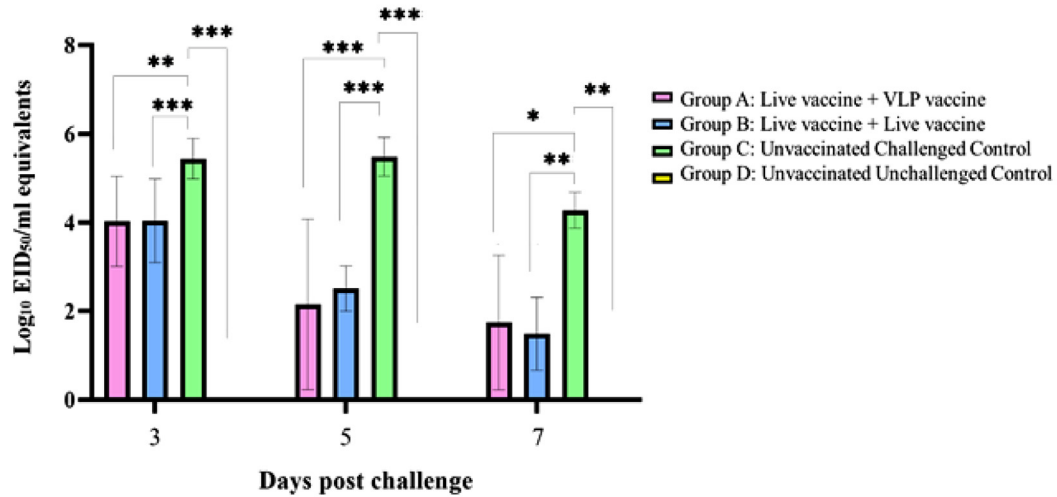


**Figure 3.** IDEXX IB ELISA antibody titers of SPF chickens vaccinated with combinations of mixed live and VLP vaccines, and challenged with live QX-like IB virus, and controls. The asterisks indicate the statistical significance \*\*\* ( $P \leq 0.001$ ); \*\*\*\* ( $P \leq 0.0001$ ). Only significant differences are indicated on the graph ( $P \leq 0.05$ ).



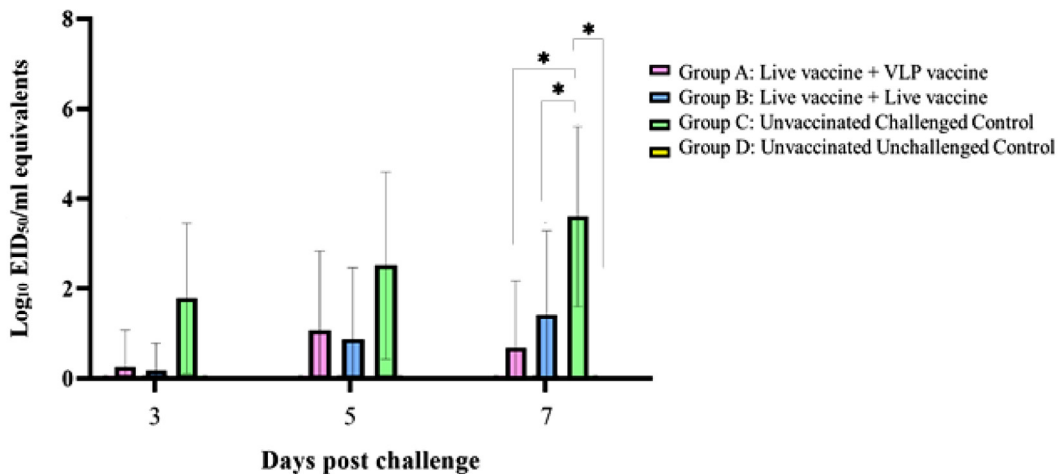
A

## Oropharyngeal Swabs



B

## Cloacal Swabs



**Figure 5.** rRT-PCR detection of QX-like IB virus in the (A) oropharyngeal and (B) cloacal swabs taken at 3-, 5-, and 7-days postchallenge. The asterisks indicate the statistical significance \* ( $P \leq 0.05$ ), \*\* ( $P \leq 0.01$ ), \*\*\* ( $P \leq 0.001$ ), \*\*\*\* ( $P \leq 0.0001$ ). Only significant differences are indicated on the graph ( $P \leq 0.05$ ).

produced a VLP batch and evaluated the efficacy of this plant-produced vaccine against homologous challenge with a QX-like virus. The use of a live-attenuated virus for the prime dose during vaccination improves the level of protection against IBV, while using inactivated vaccines offered broad-spectrum protection against multiple strains of IBV (de Wit et al., 2022). Therefore, the plant-produced VLP was applied as a booster 3 wk after vaccination with a mix of live commercial 4-91 and Ma5 IB vaccines, as simultaneous or alternate use of Ma5 and 4/91, as commonly employed in many countries, including South Africa, induces high levels of protection against heterologous IBV types such as QX (Cook et al., 1999; Terregino et al., 2008). For comparison, chickens primed and boosted with the Ma5 and 4/91 mix were included as a control group.

Prior to challenge, both vaccination regimes appeared to provide similar humoral protection as

measured by enzyme-linked immunosorbent assay (ELISA) and HI, but like all commercially available IB ELISAs, the IDEXX IB Ab ELISA is not a serotype-specific assay, and is validated to detect antibodies directed against more conserved structural IBV nucleocapsid (N) and membrane (M) proteins. The plant-produced VLP does not contain any IB proteins apart from S, therefore the commercial ELISAs probably underestimated the S-specific antibodies induced by the VLPs. The HI results, which specifically measure S-protein antibodies, were also comparable between the VLP and live vaccines, but once there is multiple exposure to IBV, as with prime-boost vaccination, the HI test displays a variety of cross-reactions with other IBV serotypes that are not homologous to the HI test antigen, creating the challenge of being unable to differentiate between varying serotypes (Cook et al., 1987; Gelb and Jackwood, 2008).

Since the QX strain 3665/11 does not induce clinical signs (e.g., respiratory signs or kidney pathology) under experimental conditions in SPF chickens (Bwala et al., 2018), we were limited in assessing the vaccine efficacy on the reduction in viral shedding and ciliary motility.

Overall, the protection against challenge was similar, with no statistically significant difference in the ability of the vaccines to reduce the magnitude of virus shedding from the oropharynx or cloaca, but boosting with the VLP vaccine did seem to reduce the duration of cloacal shedding more effectively, which in turn would aid in reducing fomite spread of IBV between flocks. The VLP vaccine booster also had an improved average protection score on the tracheal ciliary motility test, suggesting that boosting with the VLP vaccine offered better respiratory tract protection than vaccinating with the heterologous 4/91 and Ma5 vaccine mix alone. Respiratory tract protection is vital in field conditions where flocks are likely to be exposed to several pathogens. It is a limitation of this study that we used a vaccine combination for which a protectotype is available.

Ultimately, the benefit of plant-produced homologous IB VLPs over traditional whole inactivated virus vaccines is the safety, speed and scalability with which they can be produced. Bulk VLP doses can be produced in as little as 2 wk after obtaining the S gene sequence (Lai and Chen, 2012) and although there is still scope to improve the yields, in this case 1 kg of plant leaf material was sufficient to produce at least 3,352 individual 5 µg VLP vaccine doses. Like inactivated vaccines, VLP vaccines require intramuscular administration by injection, but there is the option of exploring other delivery methods such as formulated inhalable powders for mucosal immunization for the mass vaccination of flocks (Tomar et al., 2018). In conclusion, plant-produced IB VLPs have incredible potential to improve poultry health providing the ability to develop safe homologous IBV vaccines quickly and affordably, scaling them up to meet agricultural requirements of poultry industries even in lower income parts of the world, making it possible to effectively deal with the persistent global issue of emerging IBV variants.

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Ethics: The study procedure was preapproved by the Department of Agriculture, Land Reform, and Rural Development (12/11/1/1/MG), the Council for Scientific and Industrial Research Ethics committee (251/2018), and the Research and Animal Ethics Committees of the University of Pretoria (REC106-20). This research on live animals met the guidelines approved by the institutional animal care and use committee (IACUC).

Authors' contribution: K. M. S., M. O. K., C. A.: conceptualization, methodology. K. M. S., C. A., M. O. K., D. B. R. W., M. C. H., P. S.: investigation. C. A., M. O. K.: resources. C. A., K. M. S.: analysis, writing- original draft preparation. All coauthors contributed to reviewing and editing the manuscript and agree to its publication.

## DISCLOSURES

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in the present study.

## SUPPLEMENTARY MATERIALS

Supplementary material associated with this article can be found in the online version at [doi:10.1016/j.psj.2023.102953](https://doi.org/10.1016/j.psj.2023.102953).

## REFERENCES

- Andrade, L. F., P. Villegas, O. J. Fletcher, and R. Laudencia. 1982. Evaluation of ciliary movement in tracheal rings to assess immunity against infectious bronchitis virus. *Avian Dis.* 26:805–815.
- Butcher, G. D., D. P. Shapiro, and R. D. Miles. 2022. Infectious bronchitis virus: classical and variant strains. *Vet. Med.* VM127. Accessed Aug. 2023 <https://edis.ifas.ufl.edu/>.
- Bwala, D. G., P. Solomon, N. Duncan, D. B. R. Wandrag, and C. Abolnik. 2018. Assessment of *Mycoplasma gallisepticum* vaccine efficacy in a co-infection challenge model with QX-like infectious bronchitis virus. *Avian Pathol.* 47:261–270.
- Cavanagh, D. 2003. Severe acute respiratory syndrome vaccine development: experiences of vaccination against avian infectious bronchitis coronavirus. *Avian Pathol.* 32:567–582.
- Cook, J. K. A., A. J. Brown, and C. D. Bracewell. 1987. Comparison of the hemagglutination inhibition test and the serum neutralization test in tracheal organ cultures for typing infectious bronchitis virus strains. *Avian Pathol.* 16:505–511.
- Cook, J. K. A., S. J. Orbell, M. A. Woods, and M. B. Huggins. 1999. Breadth of protection of the respiratory tract provided by different live-attenuated infectious bronchitis vaccines against challenge with infectious bronchitis viruses of heterologous serotypes. *Avian Pathol.* 28:477–485.
- de Wit, J. J. S., J. K. A. Cook, and H. M. van der Heijden. 2011. Infectious bronchitis virus variants: a review of the history, current situation and control measures. *Avian Pathol.* 40:223–235.
- de Wit, J. J. S., P. de Herdt, J. K. A. Cook, M. Andreopoulou, I. Jorna, and H. C. R. Koopman. 2022. The inactivated infectious bronchitis virus (IBV) vaccine used as booster in layer hens influences the breadth of protection against challenge with IBV variants. *Avian Pathol.* 51:244–256.
- Erfanmanesh, A., A. Ghalyanchilangeroudi, D. Nikaein, H. Hosseini, and T. Mohajerfar. 2020. Evaluation of inactivated vaccine of the



- variant 2 (IS-1494 /GI-23) genotype of avian infectious bronchitis. *Comp. Immunol. Microbiol. Infect. Dis.* 71:101497.
- Gallardo, R. A. 2021. Infectious bronchitis virus variants in chickens: evolution, surveillance, control and prevention. *Aust. J. Vet. Sci.* 53:55–62.
- Gelb, J. Jr., and M. W. Jackwood. 2008. Infectious bronchitis. Pages 146–149 in *A Laboratory Manual for the Isolation, Identification and Characterization of Avian Pathogens*. L. Dufour-Zavala, D. E. Swayne, J. R. Glisson, J. E. Pearson, W. M. Reed, M. W. Jackwood and P. Woolcock, eds. 5th ed. American Association of Avian Pathologists, Kennett Square, PA.
- Hassan, M. S. H., A. Ali, S. M. Buharideen, D. Goldsmith, C. S. Coffin, S. C. Cork, F. van der Meer, M. Boulianne, and M. F. Abdul-Careem. 2021. Pathogenicity of the Canadian delmarva (DMV/1639) infectious bronchitis virus (IBV) on female reproductive tract of chickens. *Viruses* 13:2488.
- Jackwood, M. W., and J. J. de Wit. 2013. *Diseases of Poultry*. Infectious bronchitis. 13th ed. Blackwell Publishing, Ames, IA, USA, 139–160.
- Kant, A., G. Koch, D. J. van Roozelaar, J. G. Kusters, F. A. Poelwijk, and B. A. van der Zeijst. 1992. Location of antigenic sites defined by neutralizing monoclonal antibodies on the S1 avian infectious bronchitis virus glycopolyptide. *J. Gen. Virol.* 73:591–596.
- Ladman, B. S., C. R. Pope, A. F. Ziegler, T. Swieczkowski, C. J. Callahan, S. Davison, and J. Gelb Jr. 2002. Protection of chickens after live and inactivated virus vaccination against challenge with nephropathogenic infectious bronchitis virus PA/Wolgemuth/98. *Avian Dis.* 46:938–944.
- Lai, H., and Q. Chen. 2012. Bioprocessing of plant-derived virus-like particles of Norwalk virus capsid protein under current good manufacture practice regulations. *Plant Cell Rep.* 31:573–584.
- Liu, G., L. Lv, L. Yin, X. Li, D. Luo, K. Liu, C. Xue, and Y. Cao. 2013. Assembly and immunogenicity of coronavirus-like particles carrying infectious bronchitis virus M and S proteins. *Vaccine* 31:5524–5530.
- Lv, L., X. Li, G. Liu, R. Li, Q. Liu, H. Shen, W. Wang, C. Xue, and Y. Cao. 2014. Production and immunogenicity of chimeric virus-like particles containing the spike glycoprotein of infectious bronchitis virus. *J. Vet. Sci.* 15:209–216.
- Pallesen, J., N. Wang, K. S. Corbett, D. Wrapp, R. N. Kirchdoerfer, H. L. Turner, C. A. Cottrell, M. M. Becker, L. Wang, W. Shi, W. Kong, E. L. Andres, A. N. Kettenbach, M. R. Denison, J. D. Chappel, B. S. Graham, A. B. Ward, and J. S. McLellan. 2017. Immunogenicity and structures of a rationally designed prefusion MERS-CoV spike antigen. *Proc. Natl. Acad. Sci. USA* 114:E7348–E7357.
- Ruano, M., J. El-Attrache, and P. Villegas. 2000. A rapid-plate hemagglutination assay for the detection of infectious bronchitis virus. *Avian Dis.* 44:99–104.
- Sepotokele, K. M., M. M. O’Kennedy, D. B. R. Wandrag, and C. Abolnik. 2023. Optimization of infectious bronchitis virus-like particle expression in *Nicotiana benthamiana* as potential poultry vaccines. *PLoS One* 18:e0288970.
- Shirvani, E., A. Paldurai, V. K. Manoharan, B. P. Varghese, and S. K. Samal. 2018. A recombinant Newcastle disease virus (NDV) expressing S protein of infectious bronchitis virus (IBV) protects chickens against IBV and NDV. *Sci. Rep.* 8:1–14.
- Smith, T., M. M. O’Kennedy, D. B. R. Wandrag, M. Adeyemi, and C. Abolnik. 2020. Efficacy of a plant-produced virus-like particle vaccine in chickens challenged with Influenza A H6N2 virus. *Plant Biotechnol. J.* 18:502–512.
- Strasser, R., J. Stadlmann, M. Schähns, G. Stiegler, H. Quendler, L. Mach, J. Glössl, K. Weterings, M. Pabst, and H. Steinkellner. 2008. Generation of glycol-engineered *Nicotiana benthamiana* for the production of monoclonal antibodies with a homogeneous human-like N-glycan structure. *Plant Biotechnol. J.* 6:392–402.
- Terregino, C., A. Toffan, M. S. Beato, R. De Nardi, M. Vascellari, A. Meini, G. Ortali, M. Mancin, and I. Capua. 2008. Pathogenicity of a QX strain of infectious bronchitis virus in specific pathogen-free and commercial broiler chickens, and evaluation of protection induced by a vaccination programme based on the Ma5 and 4/91 serotypes. *Avian Pathol.* 37:487–493.
- Tomar, J., C. Biela, C. A. M. deHaan, P. J. M. Rottier, N. Petrovsky, H. W. Frijlink, A. Huckriede, W. L. J. Hinrichs, and B. Peeters. 2018. Passive inhalation of dry powder influenza vaccine formulations completely protects chickens against H5N1 lethal virus challenge. *Eur. J. Pharm. Biopharm.* 133:85–95.
- Ukarami, A., A. Sakurai, M. Ishikawa, M. L. Yap, Y. Flores-Garcia, Y. Haseda, T. Aoshi, F. P. Zavala, M. G. Rossman, S. Kuno, R. Ueno, and W. Akahata. 2017. Development of a novel virus-like particle vaccine platform that mimics the immature form of alphavirus. *Clin. Vacc. Immunol.* 24:1–14.
- Valastro, V., E. C. Holmes, P. Britton, A. Fusaro, M. W. Jackwood, G. Cattoli, and I. Monne. 2016. S1 gene-based phylogeny of infectious bronchitis virus: an attempt to harmonize virus classification. *Infect. Genet. Evol.* 39:349–364.
- Vaughn, D. W., and S. S. Whitehead. 2009. Dengue. *Vaccines for Biodefense and Emerging and Neglected Diseases* Elsevier, Amsterdam, the Netherlands, 285–324.
- World Bank & TAFS-Forum. 2011. World livestock disease atlas: a quantitative analysis of global animal health data (H. S. N. The World Bank, Washington, DC 20433, USA Ed.). Accessed July 2022. <https://www.tafsforum.org/papers-reports/livestock-disease-atlas>.
- WOAH, 2019. Avian infectious bronchitis. Pages 796–809 in *WOAH Terrestrial Manual*. Chapter 3.3.2. Accessed July 2022. <https://www.woah.org/en/disease/avian-infectious-bronchitis/>.
- Wu, X., X. Zhai, Y. Lai, L. Zuo, Y. Zhang, X. Mei, R. Xiang, Z. Kang, L. Zhou, and H. Wang. 2019. Construction and immunogenicity of novel chimeric virus-like particles bearing antigens of infectious bronchitis virus and Newcastle disease virus. *Viruses* 11:1–13.
- Xu, P., X. Wu, H. Wang, B. Ma, M. Ding, and X. Yang. 2016. Assembly and immunogenicity of baculovirus-derived infectious bronchitis virus-like particles carrying membrane, envelope and the recombinant spike proteins. *Biotechnol. Lett.* 38:299–304.